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Analysis of Phylogeny of Agrocybe Genus Based on Nucleotide Database in NCBI

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Analysis of Phylogeny of *Agrocybe* Genus Based on Nucleotide Database in NCBI

Abstract: The study of the phylogeny of species is a basic discipline for the effective protection, development, and utilization of their germplasm resources. Since this century, the fungi of the Agrocybe genus have become the most successful class of medium and high-grade edible and medicinal fungi that have been artificially domesticated and grown on a large scale, and have high commercial value. However, there are few reports on its germplasm resources. To clarify the genetic and developmental relationship at the genus level of the Agrocybe in the world. In this paper, Based on the NCBI Nucleotide database(National Center for Biotechnology Information), taking the single gene ITS sequence as the research object, the T92 (Tamura 3-parameters) parameter model was used to calculate the genetic distance, and the Neighbor-Joining Method was used to construct the genus-level phylogenetic tree of the Agrocybe. The results showed that the phylogenetic tree constructed by 192 ITS single gene sequences of 43 species of the Agrocybe (including 31 known species and 12 undetermined species) in the NCBI Nucleotide database was roughly divided into 6 branches and belonged to different groups, and 17 undetermined species of the Agrocybe sp. was preliminarily identified. This study has a certain reference value for the genetic relationship among different species of Agrocybe all over the world, as well as the selection of the parent bank and the protection of germplasm resources of each species of the genus. Keywords: NCBI Nucleotide Database; ITS sequence; Agrocybe genus; Phylogeny

Introduction

Edible Fungi are a kind of ideal food with rich nutrition, delicious taste, and strong fitness. They are also one of the three major foods for humans. They also have high medicinal value and are recognized as high-nutrient health food. The Agrocybe genus, which was first named by French Agrocybe FAYOD in 1889 [1], belongs to the Basidiomycota, Agaricomycetes, Agaricales, and Strophariaceae according to the 《Dictionary of the Fungi》^{10th} edition[2], the type species of the Agrocybe genus is Agrocybe praecox (Fr.)Fayod[3]. Modern medicine indicates that the fungi of the Agrocybe genus are rich in B vitamins, mineral elements, and anticancer fungal polysaccharides, The inhibition rate of mouse sarcoma¹⁸⁰ and Ehrlich ascites cancer reached 80%-90%[4]. At the same time, the Agrocybe genus has become the most successful class of high-grade edible and medicinal fungi in artificial wild domestication and large-scale planting, with high commodity value, and are favored for its unique flavor, rich nutrition, and special pharmacological efficacy[5]. Edible fungi germplasm resources are the foundation of scientific research and product development. However, in recent years, due to environmental pollution, deforestation, over-picking, and other factors, The species and quantity of the Agrocybe genus are decreasing year by year, and the selection of fine strains of the Agrocybe genus in artificial cultivation is less. Therefore, strains culture preservation is of great significance to the gene preservation of the fungal germplasm resources of the Agrocybe genus, the relative stability of the genetic performance of excellent strains, as well as its scientific research and production. At

present, most of the research on the fungi of the *Agrocybe* genus focuses on the description of macroscopic characteristics, cultivation techniques, extraction of polysaccharide components, genetic analysis, and determination, etc[6-16]. There are few reports on the diversity of its germplasm resources. Dai Yucheng *et al.* reported seven species of the *Agrocybe* genus in China[17]. Jin Xin *et al.* reported three new records of the *Agrocybe* genus is generally a composite population, in which *Agrocybe salicacola* and *Agrocybe chaxingu* should belong to independent species and be classified as *Agrocybe cylindracea*[19,20]. At present, most studies on the germplasm resources of the *Agrocybe* genus focus on the observation and determination of morphological or agronomic traits, and there is a lack of molecular phylogenetic studies at the genus level. To clarify the genetic development at the genus level of *Agrocybe* sp., this study is based on the NCBI Nucleotide database (National Center for Biotechnology Information)[21], which analyzes the genetic diversity of the germplasm resources of the *Agrocybe* genus all of the world, providing a reference for in-depth research in the future.

1. Materials and Methods

1.1 Data source

The specific information on the *Agrocybe* genus was obtained using the NCBI-Taxonomy database. A total of 43 species information was retrieved, including 31 known species information and 12 pending species information. 192 ITS (Internal Transcribed Spacer, (ITS1-5.8S rRNA-ITS2) were retrieved and collected from the NCBI-Nucleotide database for analysis.

1.2 Data analysis

Carry out homologous sequence alignment with Alignment-Clustal W of the collected *Agrocybe* genus ITS sequences by MEGA-X software, and the genetic distance was calculated by the Maximum Likelihood Method[22], The Neighbor-Joining Method constructed the phylogenetic tree of each species in *Agrocybe* genus. The confidence level of the system branches was evaluated by a self-guided test, and the bootstrap support rate of each branch was set for 1000 repeated tests[23].

2. Results and analysis

2.1 Selection of statistical models

To improve the accuracy of phylogenetic tree construction, the maximum likelihood method was used in the MEGA-X software MODELS program to find the most appropriate genetic distance statistical model in Find Best DNA / Protein Models, and the default parameters were selected during the analysis. Tab. 1 showed the maximum likelihood fit (including individual and combined models) of the 24 different nucleotide substitution models after systematic parameter statistics. In the composite model, The T92 +G combination model (BIC value =40706.37467) could be considered the best description; the AICc value of the T92 + G combined models was 36962.01904, The values were also relatively low, Slightly higher than the two combined models: GTR + G (AICc value =36944.80364) and GTR+ R + I (AICc value =36946.66629), This indicated that the T92 + G combination model had the highest overall fit degree, is the optimal solution out of the 24 substitution models. In the single model, the T92 model with BIC value =42122.97704 and AICc value =38388.3660, the lowest score and highest fit in the six single

models. Because the MEGA-X version does not provide the phylogenetic tree construction of the combined model, the T92 model with the lowest BIC value was finally selected to build the phylogenetic tree at the genus level of the *Agrocybe* genus.

Model	parameter	Bayesian Information Criterion (BIC)	Akaike Information Criterion, corrected (AICc)	Maximum Likelihood value(lnL)	Gamma distribution
T92+G	384	40706.37467	36962.01904	-18095.84777	0.861605897
HKY+G	386	40729.80454	36965.95923	-18095.80572	0.86051765
T92+G+I	385	40730.10236	36976.00188	-18101.83313	0.858041331
K2+G	383	40735.96337	37001.35262	-18116.5206	0.866323412
GTR+G	390	40747.62794	36944.80364	-18081.20347	0.871034489
TN93+G	387	40748.08624	36974.49613	-18099.06809	0.854771303
HKY+G+I	387	40753.86646	36980.27635	-18101.9582	0.853004327
GTR+G+I	391	40759.23526	36946.66629	-18081.12864	0.870953968
K2+G+I	384	40760.4191	37016.06347	-18122.86998	0.862946708
TN93+G+I	388	40773.34096	36990.00609	-18105.81696	0.84868148
JC+G	382	41011.34112	37286.47530	-18260.08797	0.880813161
JC+G+I	383	41023.14393	37288.53319	-18260.11089	0.880844977
T92+I	384	41788.70345	38044.34782	-18637.01216	n/a
K2+I	383	41807.83225	38073.22151	-18652.45505	n/a
HKY+I	386	41813.60269	38049.75738	-18637.7048	n/a
TN93+I	387	41836.52594	38062.93584	-18643.28794	n/a
GTR+I	390	41842.90036	38040.07607	-18628.83968	n/a
JC+I	382	42089.27711	38364.41129	-18799.05597	n/a
T92	383	42122.97704	38388.36630	-18810.02744	n/a
K2	382	42147.39128	38422.52546	-18828.11305	n/a
HKY	385	42148.81287	38394.71238	-18811.18838	n/a
TN93	386	42160.49446	38396.64914	-18811.15068	n/a
GTR	389	42182.82942	38389.74982	-18804.6827	n/a
JC	381	42429.16339	38714.04252	-18974.87759	n/a

Tab.1 The maximum likelihood fit of 24 different nucleotide substitution model

2.2 Construction of the phylogenetic tree

Based on the T92 model and the neighbor-joining method, the ITS sequence phylogenetic tree of the *Agrocybe* genus was constructed (Fig.1). The clustering results showed that the entire phylogenetic tree was mainly divided into 6 major branches, covering 43 species of the *Agrocybe* genus in the NCBI-Taxonomy database (including 31 known species and 12 undetermined species), and Each branch point had a high support rate (≥ 0.572). The analysis showed that Agrocybe cylindracea (Agrocybe aegerita) and Agrocybe salicacacola were most closely related to each other, and they were clustered into one class with a topological value of 0.989, which could be classified as the Sect. Aporus, the clustering results are consistent with the results of Zhang Jinxia et al[19]. Secondly, *Agrocybe erebia* with different accession numbers was clustered into a single group, with a clear phylogenetic relationship, and together with *Agrocybe brunneola*, they were grouped into the Sect. Velatae. Thirdly, *Agrocybe pediades*, and their ring-shaped variety

Agrocybe pediades, and *Agrocybe pediades* are grouped as a whole, and they are collectively classified as Sect. Pediades. In addition, *Agrocybe dura*, *Agrocybe paludosa*, and *Agrocybe praecox* are all classified as Sect. Agrocybe. It was worth noting that the accession number MN007012.1 *Agrocybe parasitica* was relatively rare and was clustered with *Agrocybe cylindracea* (*Agrocybe aegerita*) and *Agrocybe chaxingu*, and its topological value = 0.501, which is worthy of MN007012.1 *Agrocybe parasitica* specific genetic traits to carry out in-depth research.

Among the 192 ITS single gene sequence information retrieved, 17 unknown species information were to be determined (marked on the red background). Among them, the accession number KY744153.1 (Agrocybe sp. TENN 068499), MK634587.1 (Agrocybe sp. voucher JLF3250), FJ235156.1 (Agrocybe sp. SOC1251), MK607570.1 (Agrocybe sp. voucher Mushroom Observer* 303171), MK634584.1 (Agrocybe sp. voucher JLF1690), MK634586.1 (Agrocybe sp. voucher JLF1780), MK999930.1 (Agrocybe sp. isolate Mushroom Observer.org/367 657) and MK606110.1 (Agrocybe sp. voucher 46134285) 8 undetermined species were clustered with the type-species Agrocybe praecox, and there is no branch point because they belong to parallel species. Accession numbers MK018891.1 (Agrocybe sp. isolate OTU1128), MK634585.1 (Agrocybe sp. voucher JLF1775), MK627487.1 (Agrocybe sp. voucher 151 A07), and KR673463.1 (Agrocybe sp. KA12-0412) 4 pending species were clustered with Agrocybe pediades, and also belonged to parallel species. Accession numbers KF702395.1 (Agrocybe sp. ql-5), KF702393.1 (Agrocybe sp. ql-7), and KF702390.1 (Agrocybe sp. ql-10) 3 undetermined species were clustered together with Agrocybe tuberosa, Topology value ≥ 0.661 . The accession number JX135083.1 (Uncultured Agrocybe clone B20) was clustered with Agrocybe putaminum, and the topological value = 0.9343, which has a very high similarity. The accession number JN684794.1 (Agrocybe sp. LE 11405) was clustered with Agrocybe pusiola, the topology value=0.865, and the similarity was also very high.

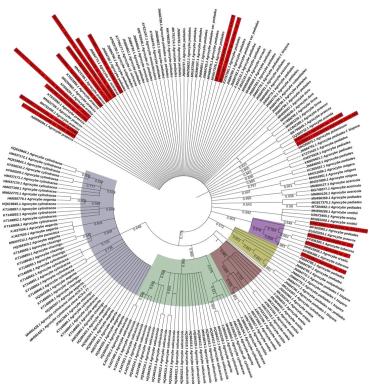


Fig.1 The phylogenetic tree of the *Agrocybe* genus based on NCBI-ITS sequence (Bootstrap, Repeat=1000)

3. Conclusions

In this study, the ITS single gene sequences of each species of the Agrocybe genus were analyzed at the Nucleotide database of the National Center for Biological Information in the United States, and the genetic distance was calculated using the T92 (Tamura 3-parametere) model. Neighbor-Joining Method constructed a phylogenetic tree of the Agrocybe genus. The results showed that the phylogenetic tree constructed by 192 ITS single gene sequences collected from the *Agrocybe* genus was roughly divided into 6 branches and belonged to different groups. Specific information is as follows: 8 pending species with accession numbers KY744153.1, MK634587.1, FJ235156.1, MK607570.1, MK634584.1, MK634586.1, MK999930.1, and MK606110.1 were classified in parallel with the type species Agrocybe praecox. The 4 pending species with accession numbers MK018891.1, MK634585.1, MK627487.1, and KR673463.1 also belonged to the parallel classification with A. pediades. The 3 undetermined species with accession numbers KF702395.1, KF702393.1, and KF702390.1 had a topological value of 0.661 with Agrocybe tuberosa, which was highly reliable. The topological value of accession number JX135083.1 and Agrocybe putaminum reached 0.943, and the similarity was extremely high, which could be regarded as the same species. The similarity of the JN684794.1 log-in numbers and Agrocybe pusiola similarity is also extremely high, with a topological value of 0.865, which can also be considered the same species. The accession number JN684794.1 was also very similar to Agrocybe pusiola, with a topological value of 0.865, which could also be regarded as the same species. In addition, special attention was paid to the accession number MN007012.1 Agrocybe parasitica, which had not yet been found in China. It was clustered with Agrocybe cylindracea (Agrocybe aegerita) and Agrocybe chaxingu. Topologically value reached 0.501, which belonged to a category with high reliability, and it was worthy of scholars to continue to carry out in-depth research on its specific genetic traits.

In 1994, Lu Chengying & Li Jianzong isolated a fungus of the Agrocybe genus in Zhangjiajie National Forest Park in Hunan Province, China, which was identified as *Agrocybe farinacea* Hongo by morphological characteristics, which was the newly recorded species in China at that time, specimen No. MHJSU940141, stored in the Jishou University Fungal Herbarium (MHJSU)[24]. In 2012, Jin Xin and Tuliguer conducted a detailed systematic study on the species taxonomy of the domestic Agrocybe, and counted three new recorded species in China, namely *Agrocybe elatella, Agrocybe brunneola,* and *Agrocybe pediades* var. cinctula Nauta, specimens of them were preserved in Jilin Agricultural University Herbarium (HMJAU)[25]. However, regarding the research of Chinese scholars on *Agrocybe farinacea, Agrocybe elatella,* and *Agrocybe brunneola*, the author did not find any information records of these three species in the NCBI-Taxonomy retrieval system, this work should be improved in future research to enrich the NCBI strain resource bank.

For a long time, the taxonomy of the *Agrocybe* genus has been chaotic, and there is no unified classification standard. The classification research of this genus at home and abroad mainly includes traditional morphology, molecular systematics, and interspecies isolation-mating test. Modern molecular phylogenetic classification has become a powerful method for the study of fungal species classification, and a variety of molecular marker techniques have emerged[26,27]. Existing molecular phylogenetic studies of the *Agrocybe* genus mainly focus on phylogenetic studies of a single gene or two gene segments, but cannot fully reflect its phylogenetic relationship. The construction of the phylogenetic tree of fungal multi-gene fragments has been adopted by the

majority of scholars, which is more conducive to the correct definition of difficult species or similar species. However, the polygenic molecular phylogeny of the *Agrocybe* genus has not been reported. Therefore, polygenic molecular phylogeny is one of the research hotspots in the molecular phylogeny of the genus Agrocybe. Due to environmental changes caused by geographical factors, the *Agrocybe* genus is unstable in morphology, and physiological and biochemical characteristics, and is prone to changes. At this time, mating experiments should be used to verify whether they produce reproductive isolation.

The phylogenetic study of species is the basic subject of biodiversity. The richer the genetic variation of a species, the stronger its survival adaptability and the greater its evolutionary potential. During its long-term natural evolution and artificial domestication, macrofungi have formed different ecological species to adapt to the ecological environment of different regions, and have relatively distinct genetic differentiation. This study has a certain reference value for the genetic relationship among different species of the *Agrocybe* genus, as well as the selection of the parent bank of each species and the protection of germplasm resources.

In recent years, with the rapid development of molecular sequencing technology, the Genealogical Concordance Phylogenetic Species Recognition (GCPSR) species identification system has become the mainstream and hot spot in the field of biodiversity research, and there have been many research reports[28,29]. Through the rapid identification of more gene families, strains morphology, and reproductive mode, this technology can help humans understand the relatedness between species more effectively.

Conflict of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

Acknowledgments

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Reference

[1]Agrocybe FAYOD. Annls sci nat, Bot., sér[M]. Fortin, Masson et Cie, Paris, 1889, 1-523.

[2]Kirk PM, Cannon PF, Minter DW, et al. Dictionary of the fungi[M]. 10th edition. CAB International, Wallingford, 2008, 1-771.

[3]Singer, R. The Agaricales in modern taxonomy [M]. 4th edition. Koeltz Scientific Books, Koengstein. 1986, 1-981.

[4]Yuan Liang, Dengxue Lu, Sen Wang, Yuhui Zhao, Shenghan Gao, Rongbing Han, Jun Yu, Weili Zheng, Jianing Geng, Songnian Hu. Genome Assembly and Pathway Analysis of Edible Mushroom *Agrocybe cylindracea*[J]. Genomics, Proteomics & Bioinformatics, 2020,18(03):341-351.

[5]Yang Yueming, Li Meiliang, Li Yinliang.Cultivation techniques of Tea tree mushroom [M].Beijing: Jindun Publishing House, 2011:15-18.(in Chinese)

[6]Meinhardt F., Leslie John F., Mating types of Agrocybe aegerita[J]. Curr Genet, 1982(5)165~68.

[7]Zhang Shuting, Lin Fangcan. Genetics and Breeding of Mushrooms [M]. Beijing: China Agricultural Press. 1997.65~119.(in Chinese)

[8]Lin Ling. Research on the application and supporting cultivation technology of *Agrocybe* spp. liquid strain[J]. Edible and Medicinal Mushrooms, 2020, 28(05):340-343. (in Chinese)

[9]Liang-Yan Liu, Zheng-Hui Li, Ze-Jun Dong, Xing-Yao Li, Jia Su, Yan Li, Ji-Kai Liu. Two novel fomannosane-type sesquiterpenoids from the culture of the basidiomycete *Agrocybe salicacola*[J]. Natural Products and Bioprospecting, 2012, 2(3).

[10]Fushimi, K., Anzai, K., Tokuyama, S., Kiriiwa, Y., Matsumoto, N., Sekiya, A., Hashizume, D., Nagasawa, K., Hirai, H., Kawagishi, H.. Agrocybynes A-E from the culture broth of *Agrocybe praecox*[J]. Tetrahedron, 2012, 68(4).

[11]Yong Tianqiao, Chen Shaodan, Xie Yizhen, Shuai Ou, Li Xiangmin, Chen Diling, Su Jiyan, Jiao Chunwei, Liang Yalei. Hypouricemic Effects of Extracts From *Agrocybe aegerita* on Hyperuricemia Mice and Virtual Prediction of Bioactives by Molecular Docking.[J]. Frontiers in pharmacology, 2018, 9.

[12]Shaoling Lin, Lai Tsz Ching, Kalung Lam, Peter C.K. Cheung. Anti-angiogenic effect of water extract from the fruiting body of Agrocybe aegerita[J]. LWT-Food Science and Technology, 2017, 75.

[13] Jie Jen Lee, Joen Rong Sheu, Woan Ching Jan, Chien Liang Liu. Water extracts of dietary mushrooms, *Agrocybe aegerita*, and *Hypsizigus mamoreus*, inhibit antigen expression of human hepatitis B virus[J]. Journal of Medicinal Plants Research, 2014, 8(5).

[14]Wang Hongxiu, Sun Peng, An Ying, Hu Jia, Chen Xutao, Li Jing, Wei Yunhui. Analysis of genetic relationship and evaluation of agronomic characters of 18 strains of Agrocybe spp.[J].Chinese Agricultural Science Bulletin, 2022, 38(01):23-31. (in Chinese)

[15]Bao Dapeng, Wang Nan, Chen Mingjie, Tan Qi, Pan Yingjie. Analysis of genetic diversity of Agrocybe aegerita strains using ARDRA and RAPD[J]. Shanghai Agricultural Journal, 2001(01):18-22.

[16]Barroso G, Labarère J. Genetic evidence for nonrandom sorting of mitochondria in the basidiomycete *Agrocybe aegerita*.[J]. Applied and environmental microbiology, 1997, 63(12).

[17]Dai Yucheng, Zhou Liwei, Yang Zhuliang, Wen'an, Tuligu, Li Taihui.Chinese Edible Fungi List [J].Journal of Bacteria, 2010,29 (01): 1-21. (in Chinese)

[18]Jin Xin, Turigur.Three new record species of Chinese Tiantou mushroom [J].Journal of Bacteria, 2012,31 (05): 795-799. (in Chinese)

[19]Zhang Jinxia, Zhao Yongchang. Germplasm Resources of edible fungi [M]. Beijing: Science Press, 2017. 607-620. (in Chinese)

[20]Jin Xin.Taxonomic study of several genera of Psilocystinidae in China [D]. Changchun: Jilin Agricultural University, 2012. (in Chinese)

[21] National Center for Biotechnology Information [DB/OL].http://www.ncbi.nlm.nih.gov/.20211120.

[22]Zhang Lili, Feng Ping, Guo Ning.An improved algorithm for evolutionary tree construction [J].Computer and Modernization, 2012,19 (02): 22-25.(in Chinese)

[23]Zhu Wen. Evolutionary Tree Construction Methods Based on the Distance Matrix Study [D]. Changsha: Hunan University, 2010.(in Chinese)

[24]Lu Chengying, Li Jianzong. Report of a newly recorded species of the Chinese field mushroom genus (Agrocybe) [J]. Journal of Jishou University (Natural Science Edition), 1994,15 (02): 85-86. (in Chinese)

[25]Jin Xin, Tuli Gu'er. Three new recorded species of the genus *T. chinensis* in China[J]. Journal of Mycology, 2012, 31(05): 795-799. (in Chinese)

[26]Petrželová Irena, Sochor Michal.How useful is the current species recognition concept for the determination of true morels? Insights from the Czech Republic.[J].MycoKeys, 2019,52,17-43.

[27]Fang Liu, Mei Wang, Ulrike Damm, et al . Species boundaries in plant pathogenic fungi: a Colletotrichum case study[J].BMC Evolutionary Biology , 2016,16(1),81-95.

[28]Matthew H. Laurence, Brett A. Summerell, Lester W. Burgess, Edward C.Y. Liew. Genealogical concordance phylogenetic species recognition in the *Fusarium oxysporum* species complex[J]. Fungal Biology,2014,118(4).

[29]Agathe Vialle, Nicolas Feau, Pascal Frey, Louis Bernier, Richard C. Hamelin. Phylogenetic species recognition reveals host-specific lineages among poplar rust fungi[J]. Molecular Phylogenetics and Evolution,2013,66(3).